

Detection of Cattle Introgression in Bison

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The existence of cattle introgression in bison was first discovered by accident while constructing the phylogeny of North American bison of the Custer State Park bison herd in South Dakota. Using mitochondrial DNA to construct their phylogenetic tree, a bovine (*Bos taurus*) mitochondrial haplotype (mitochondria1 genotype) was found within the bison population [1]. This started the entire cattle introgression issue over 20 years ago.

Detection of cattle introgressions in bison is based on the same basic principles and guidelines as used in parentage and ancestral testing in humans using molecular phylogenetic methods [2, 3]. The existence of genetic mapping databases for a host animal species provides for the identification of specific markers allowing for accurate and reliable determinations of ancestry, including introgressions.

There are 2 types of tests currently being used to detect introgression in bison: mitochondrial (mtDNA) and nuclear (nDNA) tests. Without going into detailed genetics, there are some basic understandings necessary. Every living animal has DNA, RNA, and proteins. In general, closely related animals have a high degree of agreement in their DNA (i.e., their DNA is very similar), while the DNA of animals distantly related show patterns of dissimilarity and divergence (disagreement). There are basically 2 types of DNA in mammals, the nuclear DNA found within the center of a cell (nucleus) and mitochondrial found within small organelles known as mitochondria [4].

Mitochondrial DNA (often abbreviated mtDNA) is a small piece of circular DNA found within small organelles known as mitochondria inside every cell that convert chemical energy from food into a form that cells can use, adenosine triphosphate (ATP) [5]. Mitochondrial DNA is maternally inherited (passed from the mother to her offspring) providing the ability to trace maternal lineage (matrilineality), making mtDNA the mainstay of phylogenetics (evolutionary history and relationships) and evolutionary biology. It also permits an examination of the relatedness of populations, and thus has become important in biological anthropology (biological development) and biogeography (distribution of species and ecosystems in geographic space).

Mitochondrial DNA testing is particularly important in bison because, since only crosses between bison bulls and domestic cows are successful, virtually all F1 offspring are females, and nearly all fertile backcrosses to bison are also females [6-8]. As such there is a disproportionate incidence of bovine-type mtDNA in bison [9]. The presence of bovine-type mtDNA in bison is clear evidence of cattle introgression and a hybrid animal.

However, mtDNA data alone are often hard to distinguish between closely related species to any large degree and does not provide any information on paternal (father)

HIGHLIGHTS

- There are 2 types of tests available, nuclear or autosomal DNA (nDNA) and mitochondrial DNA (mtDNA) tests.
- Both nDNA and mtDNA introgression tests need to be performed for greatest probability of detecting introgression.
- These tests identify specific markers associated with modern-day cattle and modern-day introgression and are not related to natural or ancient evolutionary processes.
- These tests detect the presence of cattle markers and not functional cattle genes.
- These markers are not a measure of the amount of cattle genes but a marker that introgression with modern-day cattle has occurred.
- Many bison have cattle genes in the absence of nDNA and mtDNA markers.

lineage (patrilineality). For instance, if a cattle bull mated with a female bison, the hybrid would be pure bison based on mtDNA analyses alone (mtDNA is based only on the mother) [5, 8]. Because of these deficiencies, other methods of analysis must also be used. This is why nuclear (autosomal) DNA tests need to be performed in addition to mtDNA testing.

Nuclear or autosomal DNA tests (nDNA) currently employed relies on the detection of differences within microsatellites of nuclear DNA [10, 11]. Understanding nDNA based tests requires a brief explanation of DNA structure. The DNA molecule is composed of 4 different chemicals known as nucleotides: adenine (A), guanine (G), cytosine (C), and thymine (T) arranged in a linear fashion [4]. The sequence of these four nucleotides at specific sites, or genes, along the chromosomes is what determines the genetic code for each individual animal. To create the double strand, G associates with C (G-C) and A associated with T (A-T). There are also specific sequences of bases repeated in a tandem fashion referred to as microsatellite DNA markers.

Microsatellites, also known as short tandem repeats or simple sequence repeats, are pieces of nuclear DNA in which small sequences (pieces) of DNA are repeated, typically 5–50 times [10]. The genomes of mammalian species may contain thousands of these microsatellite markers and they may occur at thousands of locations within an animal's genome. Because they do not code for anything (often referred to as junk DNA), they have a higher mutation rate than most other areas of DNA, leading to high genetic variability and specificity. A simple example of a microsatellite would be the sequence "...TCAGGTCTAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGTGCTTAT..." in which the sequence AG is repeated 11 times. A difference would be created by the substitution of one or more of the repeats with another nucleotide such as "...TCAGGTCTAGAGAGATAGAGAGAGAGAGAGAGAGAGTGCTTAT...". These changes are also known as single nucleotide polymorphisms or SNP (pronounced "snip") [12]. Differences may also be reflected in the number of repeats with some species having more or less.

This variability (diversity) and specificity of microsatellites make them ideal for a variety of investigations including human and wildlife forensics, parental analysis, degrees of relatedness, evidence of population bottlenecks, gene flow between populations, genetic fingerprinting, in addition to their application in cancer and other disease diagnoses [10]. Because of the variability and specificity of microsatellites, selecting and examining appropriate microsatellites can provide the ability to differentiate closely related animals and even identify individual animals, e.g., "fingerprinting". As such, all animals can be separated and identified with microsatellite nuclear DNA examination.

Specific microsatellites sequences have been identified that are unique to a host of mammalian species including modern day cattle and bison [11, 13-15]. Specific cattle associated microsatellites should not be present in bison and their detection is an indication of introgression. There are currently 15-25 microsatellites in bison that are currently used to identify cattle introgression. If you have tests performed through the National Bison Association, they will only examine 15 microsatellite markers; if you go directly to the laboratory at the University of California-Davis, you will get the extended examination with 25 microsatellites examined.

The detection of any cattle-associated microsatellites or bovine-type mtDNA is clear evidence of past cattle introgression and the presence of cattle genes.

Interpreting the results is a little more difficult than understanding the tests.

Results are often discussed as cattle genes or alleles, which are a variant form of a gene [16]; however, a gene encodes a functional product which microsatellites and other non-coding sequences do not. Rather, the presence of a cattle microsatellite or bovine mtDNA denotes the detection of a

marker of cattle introgression and does not reflect the amount of cattle-associated genes that may be present. Thus, an animal with a single cattle microsatellite marker could have more cattle genes than a bison with 3 cattle-associated microsatellites.

Efforts to suggest a percentage of cattle “genes” in bison based on microsatellite markers are totally erroneous and a misinterpretation of the data. As noted above, these microsatellites are not genes or gene alleles, and although they MAY be a reflection of the amount of cattle genes present, they are only a marker for the existence of past introgression. It is erroneous to claim that the amount of cattle DNA in bison is low, ranging from 0.56 to 1.8% of the total nuclear DNA [17, 18]; these figures are the calculated estimate of the amount of these microsatellite markers and not the amount of functional cattle genes. If some humans can contain as much as 4-6% Denisovans DNA after 70,000-100,000 years, it is erroneous to suggest that the bovine genes can be reduced to >1% in bison after only 100+ years and 15-20 generations. While the cattle markers may represent <1% of the bison genome, it is not an indication of the amount of cattle genes within the animal. It is markers that are being detected, not cattle genes.

In addition, the lack of any cattle-associated markers does not necessarily mean that there was no introgression, only that it was not detected by the methods currently used. There are many examples whereby an animal with known cattle introgression in its history (phylogeny) has no demonstrable introgression by current methods. Take a bison bull with cattle-type mtDNA but no nuclear markers (there are many of these animals). This bull unequivocally had a bovine cow as a distant grandmother in his past and unequivocally has cattle genes, just doesn’t have any of the known and verified nuclear markers. Since mtDNA is not passed by the male, this bull will pass his cattle genes to all his progeny but no nuclear cattle-associated microsatellites or mtDNA, i.e., his offspring will test as pure bison when they are not. Another example of cattle-bison introgression without mtDNA or nDNA markers includes the “white buffalos” that contain the Charolais silver locus protein homolog (SILV) gene often with normal bison-type mtDNA and no nuclear cattle markers.

These are just a few of the known cattle genes, as opposed to markers, that are not readily detected by current methods. Further examination of the bison genome will shed light on these markers and the presence of genuine cattle genes, or their associated regions, and uncover the true extent of cattle introgression in modern-day bison.

Efforts are currently in progress to define the bison genome and identify the actual cattle genes present. Using massive parallel sequencing (also called next-generation sequencing or NGS [19]), investigators at Texas A & M are attempting to sequence the bison genome. This may define the actual amount of cattle genes in introgressed bison once the complete sequence of the “pure” bison genome is known and corroborated by others. However, while a great deal of valuable information will be derived from these studies, they are not without difficulties. The technology of NGS produces short reads (50-400 nucleotides at a time) and relies on

HIGHLIGHTS

- The DNA introgression tests do not detect cattle genes but only identify that cattle introgression has occurred.
- It is erroneous to suggest that these markers reflect the amount of modern-day cattle genes present.
- Bison with evidence of cattle introgression are hybrid animals by definition.
- The absence of nDNA or mtDNA cattle markers is not a guarantee that the animal is not introgressed.
- There is no data supporting the suggestions that the amount of cattle genes in bison is low.

computer algorithms to assemble these short reads into longer lengths [19]. This assembly is not without error. Determining the true bison sequence will require sequencing more than just a handful of animals and will require independent corroboration. Although NGS is becoming more affordable and super-computers for sequence assembly are readily available, the technology is relatively expensive which hinders efforts and the number of animals that can be examined. In addition to limitations in technology, sequencing the bison genome by NGS may present other difficulties and complications.

Since we do not know the true extent of introgression, the issue of what constitutes a pure bison may come into question; can enough bison be sequenced to determine “pure” bison within a reasonable degree of certainty? It is also very possible that all bison have some evidence of cattle genes (actual genes as opposed to markers) due to cattle introgression in the late 1800’s and early 1900’s. This has its own implications. In contrast, if NGS determines that there are relatively few genetically pure bison, it could open a can of worms including the possibility that pure bison could get listed as an endangered or threatened species with wide implications to the industry [20].

It is clear that the issue of cattle-bison introgression continues to be an evolving situation that is not likely to go away in the near future.

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References

- [1] Polziehn RO, Strobeck C, Sheraton J, Beech R. Bovine mtDNA Discovered in North American Bison Populations. Conservation Biology. 1995;9:1638-43. Available at:
http://www.ozarkbisons.com/literature/genetics/polziehn_1995.pdf

- [2] Wikipedia. Molecular phylogenetics. 2018. Available at: https://en.wikipedia.org/wiki/Molecular_phylogenetics
- [3] Wikipedia. Genealogical DNA test. 2018. Available at: https://en.wikipedia.org/wiki/Genealogical_DNA_test
- [4] Wikipedia. DNA. 2018. Available at: <https://en.wikipedia.org/wiki/DNA>
- [5] Wikipedia. Mitochondrial DNA. 2018. Available at: https://en.wikipedia.org/wiki/Mitochondrial_DNA
- [6] Boyd MM. Crossing bison and cattle: First Cross Dangerous But Results are Better in Each Succeeding Generation—Hope of Taking Fur and Hump of Bison and Placing Them Upon Back of Domestic Ox. . J Heredity. 1914;5:189-97. Available at: http://www.ozarkbisons.com/literature/genetics/boyd_1914.pdf
- [7] Goodnight C. My experience with bison hybrids. J Heredity. 1914;5:197-9. Available at: http://www.ozarkbisons.com/literature/genetics/goodnight_1914.pdf
- [8] Hedrick PW. Conservation genetics and North American bison (Bison bison). The Journal of heredity. 2009;100:411-20. Available at: http://www.ozarkbisons.com/literature/genetics/hedrick_2009.pdf
- [9] Hedrick PW. Cattle ancestry in bison: explanations for higher mtDNA than autosomal ancestry. Molecular ecology. 2010;19:3328-35. Available at: http://www.ozarkbisons.com/literature/genetics/hedrick_2010.pdf
- [10] Wikipedia. Microsatellite. 2018. Available at: <https://en.wikipedia.org/wiki/Microsatellite>
- [11] Schnabel RD, Ward TJ, Derr JN. Validation of 15 microsatellites for parentage testing in North American bison, Bison bison and domestic cattle. Animal genetics. 2000;31:360-6. Available at: http://www.ozarkbisons.com/literature/genetics/schnabel_2000.pdf
- [12] Wikipedia. Single-nucleotide polymorphism. 2018. Available at: https://en.wikipedia.org/wiki/Single-nucleotide_polymorphism
- [13] Schnabel RD, Taylor JF, Derr JN. Development of a linkage map and QTL scan for growth traits in North American bison. Cytogenetic and genome research. 2003;102:59-64. Available at: http://www.ozarkbisons.com/literature/genetics/schnabel_2003.pdf
- [14] Halbert ND, Ward TJ, Schnabel RD, Taylor JF, Derr JN. Conservation genomics: disequilibrium mapping of domestic cattle chromosomal segments in North American bison populations. Molecular ecology. 2005;14:2343-62. Available at: http://www.ozarkbisons.com/literature/genetics/halbert_2005.pdf
- [15] Cronin MA, Leesburg VL. Genetic variation and differentiation in parent-descendant cattle and bison populations. Journal of animal science. 2016;94:4491-7. Available at: http://www.ozarkbisons.com/literature/genetics/cronin_2016.pdf
- [16] Wikipedia. Allele. 2018. Available at: <https://en.wikipedia.org/wiki/Allele>
- [17] Wikipedia. American bison. 2018. Available at: https://en.wikipedia.org/wiki/American_bison
- [18] Halbert ND, Gogan PJP, Heibert R, Derr JN. Where the buffalo roam: The role of history and genetics in the conservation of bison on U.S. federal lands. Park Science. 2007;24. Available at: http://www.ozarkbisons.com/literature/genetics/halbert_2007.pdf
- [19] Zhang J, Chiodini R, Badr A, Zhang G. The impact of next-generation sequencing on genomics. Journal of genetics and genomics 2011;38:95-109. Available at: http://www.ozarkbisons.com/literature/genetics/zhang_2011.pdf
- [20] Gear KO, Gear WM. Bison Genetics - The new war against bison. 2010. Available at: http://www.ozarkbisons.com/literature/genetics/gear_2010.pdf